

GADAIN, A LIGNAN FROM *JATROPHA GOSSYPIFOLIA*

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Abstract—The isolation of a new lignan, gadain, from *Jatropha gossypifolia* is reported. The structure and stereochemistry of this compound have been determined from spectral analysis, partial synthesis from jatrophan (a lignan of known absolute configuration) and from its transformation reactions.

INTRODUCTION

In continuation of our work [1] on the constituents of *Jatropha gossypifolia* L., we isolated a new lignan, gadain (1a). In this paper we report the structure of this new compound, its interesting chemistry, as well as its partial synthesis from jatrophan (2). Several reactions of jatrophan (2) are also discussed.

RESULTS AND DISCUSSION

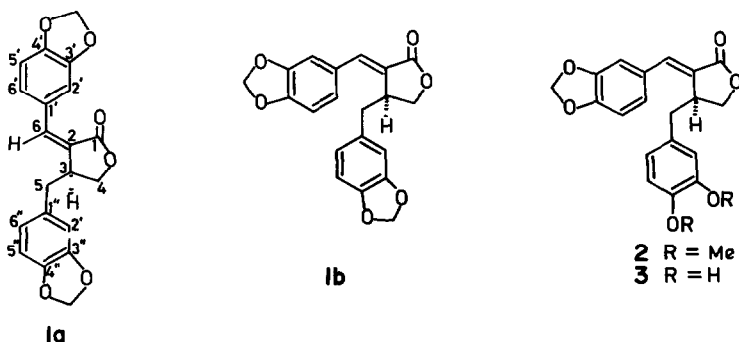
Lignan 1a, C₂₀H₁₆O₆ ([M]⁺ *m/z* 352), mp 145° (C₆H₆), [α]_D²⁵ +86° (CHCl₃), exhibited UV absorption [λ_{max}^{EtOH} nm (log ε) 337 (4.20), 293 (4.05) and 235 (4.12)] characteristic of lignans containing a dibenzylbutyrolactone skeleton with a double bond at the 2,6-position of the γ-butyrolactone ring [1-3]. The IR spectrum showed characteristic signals of an α,β-unsaturated γ-lactone (1725 cm⁻¹), olefinic double bond (1620 cm⁻¹), aromatic nucleus (1590, 1490 and 1470 cm⁻¹) and methylenedioxy group (915 cm⁻¹). The 80 MHz ¹H NMR spectrum (CDCl₃) of 1a revealed the presence of two methylenedioxy groups (δ 5.97, 2H, *s* and 5.92, 2H, *s*), one highly deshielded aromatic proton (7.71, *s*) and five more aromatic and an olefinic proton (7.09-6.57, *m*). The deshielding of one of the six aromatic protons indicated its proximity to the carbonyl group. H-3, H₂-4 and H₂-5 appeared as multiplets at *ca* δ 3.32, 4.41-4.02 and 2.89-2.76, respectively. Characteristic fragments were observed at *m/z* 352 [M]⁺, 217, 189, 135 (base peak), 77

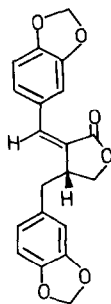
and 28 in the mass spectrum. The peaks at *m/z* 217 and 135 resulted from benzylic cleavage at the 3,5-position. The structure of gadain therefore appeared to be 1a with the deshielded H-2'.

On the basis of its 100 MHz ¹³C NMR and 300 MHz ¹H NMR spectra, the structure and stereochemistry of gadain could be unambiguously confirmed as 1a. The carbon chemical shifts were assigned from the additivity relationship as well as from a comparison with jatrophan (2) (Table 1). The values of the latter had not been reported earlier [1].

In the ¹³C NMR spectrum, 20 resolved lines were obtained. The APT experiment confirmed the presence of two >CH₂ groups, one next to oxygen (δ 69.80) and one bonded only to carbon (δ 40.73), a -CH (C-3) group (δ 44.22), two methylenedioxy carbons (δ 101.42 and 101.02) together with seven protonated *sp*² carbons [olefinic carbon C-6 at δ 140.33 and six aromatic doublets (C-2', δ 108.36, C-5', 110.34, C-6', 125.18, C-2'', 107.87, C-5'', 109.27, C-6'', 122.69)] and seven non-protonated *sp*² carbons (C-1', δ 131.37, C-1'', 127.89, C-3', 147.92, C-3'', 146.46, C-4', 148.97, C-4'', 147.55, C-2, 126.85). The δ 169.26 shift is consistent with a conjugated lactone.

While studying the 300 MHz ¹H NMR spectrum of gadain in order to confirm the disposition of H-6, we observed an interesting *cis-trans* isomerization. This conversion is possibly catalysed by the usual trace of hydrochloric acid in CDCl₃, the spectrum being run after leaving the compound in solution for 24 hr. At this stage,





1c

Table 1 ^{13}C NMR spectral data of **1a** and **2** (CDCl_3 , TMS as internal standard, δ values in ppm)

	1a	2
C-1	169.26	172.21
C-2	126.85	125.97
C-3	44.22	39.64
C-4	69.80	69.53
C-5	40.73	37.33
C-6	140.33	136.71
C-1'	131.37	130.17
C-2'	108.36	108.43
C-3'	147.92	148.80
C-4'	148.97	148.91
C-5'	110.34	112.03
C-6'	125.18	125.88
C-1''	127.89	127.91
C-2''	107.87	108.23
C-3''	146.46	147.84
C-4''	147.55	148.08
C-5''	109.27	111.33
C-6''	122.69	120.69
-OMe	—	55.60
-OCH ₂ O-	101.42, 101.02	101.51

two species were found to be present **1a** and **1b** in a ratio of 2:1. The reaction was not reversible and with time, the ratio of the peak intensities reversed. The 30% component **1b** became 60%. In fact, in keeping **1a** in hydrochloric acid (1.0 M) for 72 hr, it changed completely to **1b**, $\text{C}_{20}\text{H}_{16}\text{O}_6$ ($[\text{M}]^+$ m/z 352), mp 139° (C_6H_6). The two sets of ^1H NMR values are given in Table 2.

As the presence of the two species caused crowding in the 300 MHz ^1H NMR spectrum, the 2D HOMCOR (COSY) experiment was carried out. Both isomers exhibited the sequence $-\text{CH}_2-\text{CH}-\text{CH}_2-\text{O}-$. The aromatic region showed two ABX patterns, the minor one being more compact. Of special significance was the shift of the olefinic proton by 0.95 ppm from δ 6.59 to 7.54. The larger chemical shift is associated with the olefinic proton *cis* to the carbonyl and hence gadain must have the olefinic proton *trans* to the carbonyl, i.e. *cis* (Z) double bond [4] as in **1a**. The upfield shift of the aromatic H-2' (from δ 7.75 to 7.07) in the more stable form **1b** accompanied with the

Table 2 300 MHz ^1H NMR spectral data of **1a** and **1b** (CDCl_3 , TMS as internal standard, δ values in ppm, J in Hz)

	1a	1b
H-6	6.59, <i>d</i> , 1H $J = 1.6$	7.54, <i>d</i> , 1H $J = 1.6$
H-2'	7.75, <i>d</i> , 1H $J = 1.5$	7.07, <i>d</i> , 1H $J = 1.5$
H-5'	6.80, <i>d</i> , 1H $J = 8.0$	6.89, <i>d</i> , 1H $J = 8.0$
H-6'	7.17, <i>dd</i> , 1H $J_1 = 8.0, J_2 = 1.5$	7.11, <i>dd</i> , 1H $J_1 = 8.0, J_2 = 1.5$
H-2''	6.70, <i>d</i> , 1H $J = 1.5$	6.68, <i>d</i> , 1H $J = 1.5$
H-5''	6.66, <i>d</i> , 1H $J = 8.0$	6.64, <i>d</i> , 1H $J = 8.0$
H-6''	6.77, <i>dd</i> , 1H $J_1 = 8.0, J_2 = 1.5$	6.75, <i>dd</i> , 1H $J_1 = 8.0, J_2 = 1.5$
H _A -4	4.12, <i>dd</i> , 1H $J_1 = 10.8, J_2 = 4.8$	4.32-4.24, <i>m</i> , 2H
H _B -4	4.34, <i>dd</i> , 1H $J_1 = 10.8, J_2 = 8.4$	
H-3	3.31, <i>m</i> , 1H	3.72, <i>m</i> , 1H
H _A -5	2.82, <i>dd</i> , 1H $J_1 = 16.8, J_2 = 10.8$	2.60, <i>dd</i> , 1H $J_1 = 16.8, J_2 = 12.0$
H _B -5	2.92, <i>dd</i> , 1H $J_1 = 16.8, J_2 = 8.4$	3.02, <i>dd</i> , 1H $J_1 = 16.8, J_2 = 4.8$
-OCH ₂ O-	6.0, <i>s</i> , 2H	6.06, <i>s</i> , 2H
-OCH ₂ O-	5.96, <i>dd</i> , 2H $J_1 = 3.0, J_2 = 1.2$	5.93, <i>dd</i> , 2H $J_1 = 3.0, J_2 = 1.2$

downfield shift of the olefinic proton provided strong support for the *cis*-*trans* isomerization observed on keeping gadain in CDCl_3 .

The structure and stereochemistry of gadain received further confirmation from its synthesis from jatrophan (**2**). The latter on demethylation with boron tribromide afforded a hydroxylactone, **3**, $\text{C}_{19}\text{H}_{16}\text{O}_6$, mp 215° (C_6H_6). The presence of the hydroxyls was confirmed from the IR absorption peak at 3380 cm^{-1} and the broad two-proton signal at δ 9.65 which disappeared on deuteration. The chemical shifts for the other protons were almost similar to those of the corresponding protons in **2**. On treatment with bromochloromethane in dry acetone in the presence of anhydrous potassium carbonate, compound **3** afforded **1b**, $\text{C}_{20}\text{H}_{16}\text{O}_6$ ($[\text{M}]^+$ 352), mp 142° ($\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 1730, 1630, 1610, 1485 and 915). It was found to be identical with the product isolated earlier from hydrochloric acid isomerization of the naturally-occurring lignan, gadain. Irradiation of compound **1b** in acetone with a medium pressure mercury vapour lamp simultaneously at 313 and 336 nm for 12 hr afforded its isomer, mp 144°, $[\alpha]_{\text{D}}^{25} + 86^\circ$, which was found to be identical with the natural product, gadain (**1a**), from its physical and spectral properties. This synthesis established the 3*S*-configuration for **1a**. This stereochemistry was also supported by the comparison of its physical and spectral properties to those of (-)-isohiballactone (**1c**) [3] formed by UV irradiation of (-)-hiballactone. Gadain (**1a**) showed similar properties to (-)-isohiballactone (**1c**) (3*R*-configuration) with opposite optical rotation. Hence gadain must be *cis*(Z)-2-piperonylidene-3-piperonyl-3-*S*- γ -butyrolactone.

Both gadain (**1a**) and jatrophin (**2**) possess an α,β -unsaturated γ -lactone system and hence would be susceptible to metal hydride reduction. On reduction with both lithium aluminium hydride in tetrahydrofuran and sodium borohydride in methanol, lignans **1a** and **2** afforded the corresponding diols **4** [$C_{20}H_{20}O_6$ ($[M]^+$ m/z 356), mp 135° (C_6H_6)] and **5** [$C_{21}H_{24}O_6$ ($[M]^+$ m/z 372), mp 119° (C_6H_6)], respectively. Their IR spectra revealed the presence of hydroxyl groups (ν_{\max}^{KBr} cm^{-1} for **4** 3375, for **5** 3380) but lacked the peak absorptions due to the α,β -unsaturated γ -lactone. The 1H NMR spectrum of **4** and **5** displayed a broad signal at δ 2.27 (2H) and 2.16 (2H), respectively, which disappeared on deuteration. They underwent allylic oxidation with manganese dioxide yielding gadain (**1a**) and jatrophin (**2**), respectively.

Jatrophin (**2**) on oxidation with a mixture of 10% aqueous potassium hydroxide and 5% aqueous potassium permanganate yielded veratric acid (**6**), mp 181° (Et_2O) (lit [5] 181 – 182°) and piperonylic acid (**7**), mp 228° (Et_2O) (lit [6] 229°), while gadain (**1a**) gave only piperonylic acid (**7**), mp 228° (Et_2O).

Hydrogenation of the unsaturated lignans **1a** and **2** produced the dihydro-compounds **8** [$C_{20}H_{18}O_6$ ($[M]^+$ m/z 354), mp 132° (C_6H_6)] and **9** [$C_{21}H_{22}O_6$ ($[M]^+$ m/z 370), mp 125° (C_6H_6)], respectively. The 1H NMR spectra clearly showed the absence of H-6 in **8** and **9**.

Gadain (**1a**) underwent an interesting oxidative cyclization with DDQ in refluxing benzene, affording the naturally-occurring aryl-naphthalide lignan, justicidin E (**10**) [7], $C_{20}H_{12}O_6$ ($[M]^+$ m/z 348), mp 265° (C_6H_6) (lit [7] 265 – 269°). This obviously indicated that the naturally-occurring lignan gadain (**1a**) isomerized to the thermodynamically more stable **1b** prior to cyclization. Under similar conditions, jatrophin (**2**) afforded retrochunensin (**11**) [8], $C_{21}H_{16}O_6$ ($[M]^+$ m/z 364), mp 232° (lit [8] 234 – 236°).

Osmic acid oxidation of jatrophin (**2**) converted it to a 2,3-dibenzylbutyrolactone lignan, **12**, $C_{21}H_{22}O_8$ ($[M]^+$ m/z 402), mp 175° ($EtOH$) possessing vicinal OH-2 and OH-6 (ν_{\max}^{KBr} cm^{-1} 3560, and δ ($CDCl_3$) 2.63, 2H, *br s*, exchangeable with D_2O). Acidification of **12** with hydrochloric acid (12 M) and acetic acid (1:25) converted it into an aryltetralin lignan, **13**, $C_{21}H_{20}O_7$ ($[M]^+$ m/z 384), mp 245° ($EtOH$) (ν_{\max}^{KBr} cm^{-1} 3520, and δ ($CDCl_3$) 2.58, 1H, *br s*, exchangeable with D_2O). When **13** was refluxed with 10% Pd-C in *p*-cymene for 2 hr, it was completely aromatized to a naturally-occurring aryl-naphthalide lignan, justicidin B (**14**) [9], $C_{21}H_{16}O_6$, mp 235° ($EtOH$) (lit [9] 235 – 238°). The latter was also prepared directly from jatrophin (**2**) by NBS treatment.

The reactions are summarized in Scheme 1.

EXPERIMENTAL

Plant material. Seeds, roots and stem of *J. gossypifolia* L. were collected from Nadia District, West Bengal, India. Voucher specimens JG(se), JG(r) and JG(st) have been preserved in our laboratory.

Isolation of gadain (1a). Air-dried and finely milled stem, roots and seeds (30 kg) were exhaustively extracted with petrol (60–80°) in a Soxhlet apparatus for 72 hr. The extract was concd and chromatographed over silica gel, the column being eluted with solvents of increasing polarity. The C_6H_6 eluate afforded gadain (yield 60 mg, 0.0002%), $C_{20}H_{16}O_6$, mp 145° (C_6H_6).

Reaction of 1a with HCl. Gadain (5 mg) was dissolved in $CHCl_3$ (0.5 ml) and a drop of HCl (1.0 M) added. The mixture

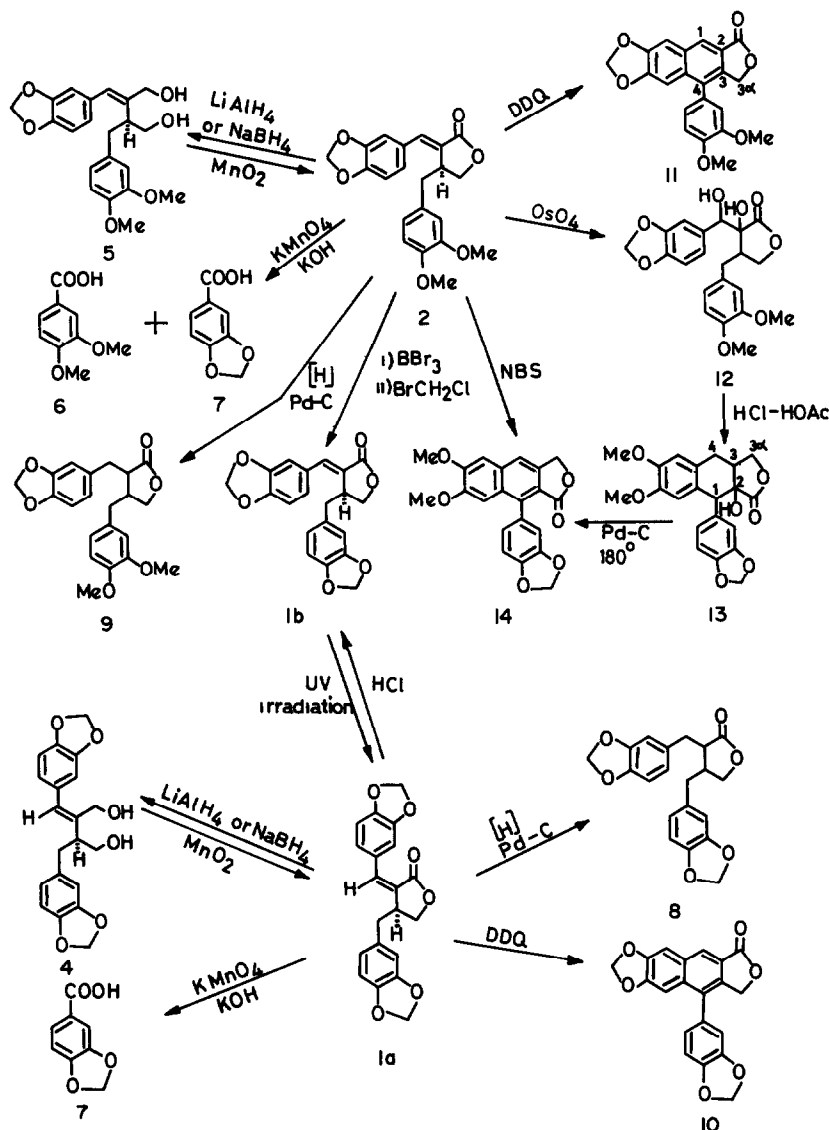
was kept at room temp for 72 hr. It was then diluted with $CHCl_3$, washed with H_2O and dried (Na_2SO_4). The conc. mass was purified by prep. TLC on silica gel using C_6H_6 -EtOAc (9:1). **1b** was obtained as white needles (3 mg), $C_{20}H_{16}O_6$, mp 139° (C_6H_6), IR ν_{\max}^{KBr} cm^{-1} 1730 (α,β -unsaturated γ -lactone), 1630 ($\gt C=C\lt$), 1610, 1485 (aromatic $\gt C=C\lt$), 915 ($-OCH_2O-$).

Synthesis of 1a (a) Demethylation of 2. To a soln of **2** (30 mg) in dry CH_2Cl_2 (20 ml) maintained at 0° to -5° , BBr_3 (1 ml) diluted with CH_2Cl_2 (10 ml) was added dropwise with vigorous stirring (2 hr) under N_2 . The soln was then allowed to stand for 12 hr at room temp. Excess BBr_3 , and the boron complexes formed in the reaction, were decomposed with ice-cold H_2O and the CH_2Cl_2 layer was washed with 2% aq. $NaHCO_3$ soln (4 \times 20 ml) and H_2O , and finally dried (Na_2SO_4). The conc. CH_2Cl_2 soln yielded the demethylated product **3** (14 mg), $C_{19}H_{16}O_6$, mp 215° (C_6H_6), IR ν_{\max}^{KBr} cm^{-1} 3380 ($-OH$), 1725 (α,β -unsaturated γ -lactone), 1625 ($\gt C=C\lt$), 1590, 1480 (aromatic $\gt C=C\lt$), 910 ($-OCH_2O-$), 1H NMR ($CDCl_3$) δ 9.65 (2H, *br s*, exchangeable with D_2O , 2-OH), 7.50 (1H, *s*, H-6), 7.0–6.5 (6H, *m*, Ar-H), 6.02 (2H, *s*, $-OCH_2O-$), 4.25 (2H, *d*, $J = 4.1$ Hz, H-4), 3.61 (1H, *m*, H-3), 3.03–2.64 (2H, *m*, H-2, 5), MS m/z 340 [$M]^+$, 217, 189, 159, 123 (base peak), 77 and 28. (b) **Alkylation of 3 with BrCH₂Cl.** The demethylated product **3** (10 mg) was alkylated with $BrCH_2Cl$ (0.2 ml) in refluxing dry Me_2CO (20 ml) containing dry K_2CO_3 (200 mg). The filtrate on concn afforded a gummy residue which was crystallized from C_6H_6 to yield **1b** (5 mg), $C_{20}H_{16}O_6$, mp 142° . (c) **UV irradiation of 1b.** **1b** (4 mg) was dissolved in Me_2CO (5 ml) and irradiated with UV light (313 and 336 nm) with a medium pressure Hg vapour lamp for 12 hr. The light yellow soln showed two spots on TLC [R_f 0.49 and 0.53, silica gel, C_6H_6 -EtOAc (9:1)]. The more polar spot corresponded to **1b** and the less polar spot to gadain (**1a**). The compounds were separated by prep. TLC on silica gel using C_6H_6 -EtOAc (9:1), and crystallization of the product from the band of higher R_f gave gadain (**1a**, 2 mg), mp 144° (C_6H_6), $[\alpha]_D^{25} + 86^\circ$ ($CHCl_3$).

$LiAlH_4$ reduction of 2. A soln of **2** (30 mg) in dry THF (50 ml) was added dropwise to a slurry of $LiAlH_4$ (20 mg) in THF at 0° under dry conditions with vigorous stirring (6 hr). Excess reagent was decomposed with EtOAc and ice-cold H_2O . The soln was filtered and the residue washed with hot EtOAc. The EtOAc soln was concd to yield the diol **5** (19 mg), $C_{21}H_{24}O_6$, mp 119° (C_6H_6), UV λ_{\max}^{EtOH} nm 257 (log ϵ 3.82), IR ν_{\max}^{KBr} cm^{-1} 3380 ($-OH$), 1595, 1485 (aromatic residue and $\gt C=C\lt$), 920 ($-OCH_2O-$), 1H NMR ($CDCl_3$) δ 7.42 (1H, *s*, H-6), 6.64–6.46 (6H, *m*, Ar-H), 5.89 (2H, *s*, $-OCH_2O-$), 4.42–4.04 (4H, *m*, H-2-1 and H-2-4), 3.81 and 3.71 (each 3H, *s*, 2-OMe), 3.50 (1H, *m*, H-3), 2.72–2.40 (2H, *m*, H-2, 5) and 2.27 (2H, *br s*, exchangeable with D_2O , 2-OH), MS m/z 372 [$M]^+$, 354, 221, 203, 151 (base peak), 77, 28.

$NaBH_4$ reduction of 2. A MeOH suspension of **2** (30 mg) was cooled in ice and treated with $NaBH_4$ (20 mg) in small portions. The reaction was monitored by TLC. At the end of the reaction, MeOH was removed and H_2O (20 ml) added to the reaction mixture. The soln was kept overnight and finally extracted with $CHCl_3$. The $CHCl_3$ soln was concd when the diol **5** (16 mg), $C_{21}H_{24}O_6$, mp 119° (C_6H_6), was obtained. It was found to be identical to the $LiAlH_4$ reduction product of **2** from mmp, co-TLC and superimposable IR spectra.

The above two reactions were repeated with **1a** (10 mg). Both produced the diol (**4**), $C_{20}H_{20}O_6$, mp 135° (C_6H_6), IR ν_{\max}^{KBr} cm^{-1} 3375 ($-OH$), 1600, 1490 (aromatic residue and $\gt C=C\lt$), 920 ($-OCH_2O-$), 1H NMR ($CDCl_3$) δ 7.61 (1H, *s*, Ar-H), 6.82–6.54 (6H, *m*, Ar-H and H-6), 5.97 and 5.90 (each 2H, *s*, 2- OCH_2O-), 4.48–4.02 (4H, *m*, H-2-1 and H-2-4), 3.50 (1H, *m*, H-3), 2.82–2.46 (2H, *m*, H-2, 5) and 2.16 (2H, *br s*, exchangeable with D_2O , 2-OH),



Scheme 1 Reactions of jatrophane (2) and gadane (1a)

MS m/z 356 $[M]^+$, 338, 221, 203, 135 (base peak), 77, 28

Oxidation of diols 4 and 5 with MnO_2 4 and 5 were oxidized with MnO_2 to gadane (1a) and jatrophane (2), respectively, according to the following procedure. The diol (4 or 5, 5 mg) in dry Me_2CO (20 ml) was stirred for 20 hr with freshly precipitated MnO_2 (30 mg). The mixture was filtered and the filtrate evapd to a gummy material which was crystallized from C_6H_6 . The products were identified by mmp, co-TLC and superimposable IR spectra with authentic 1a and 2. Yields for the oxidations of 4 and 5 were 3.5 and 4.0 mg, respectively.

Oxidations of 2 and 1a with $KMnO_4$ A mixture of 2 (30 mg), 10% aq. KOH (10 ml) and 5% aq. $KMnO_4$ (20 ml) was refluxed at 100° (2 hr). The reaction mixture was cooled and decolorized with Na_2SO_3 soln. The soln was filtered, acidified with H_2SO_4 (2 N) and extracted with Et_2O (5×50 ml). The extract was dried and concd. The mixture was subjected to fractional crystallization from Et_2O when two products, veratric acid (6, 4.5 mg), mp 181° (Et_2O) (lit [5] 181 – 182°) and piperonylic acid (7, 12.5 mg), mp 228° (Et_2O) (lit [6] 229°) were obtained. Both compounds

were identified by mmp, co-TLC and superimposable IR spectra with authentic samples.

Gadane (1a, 10 mg) was also oxidized with alkaline $KMnO_4$ soln following the above procedure to furnish piperonylic acid (7, 6.0 mg), mp 228° (Et_2O) (lit [6] 229°).

Hydrogenation of 2 and 1a Jatrophane (2, 20 mg) was dissolved in $EtOH$ (20 ml) and the soln saturated with H_2 over 10% Pd-C for 4 hr. The filtrate on concn afforded a gummy residue which crystallized from C_6H_6 as a white solid, 9 (16 mg), $C_{21}H_{22}O_6$, mp 125° (C_6H_6), IR $\nu_{max}^{KBr} cm^{-1}$ 1775 (γ -lactone), 1598, 1518, 1490 (aromatic $>C=C<$), 1190 ($-OMe$) and 930 ($-OCH_2O-$), 1H NMR ($CDCl_3$) δ 7.06–6.43 (6H, *m*, Ar-H), 6.02 (2H, *s*, $-OCH_2O-$), 4.12 (2H, *br s*, H_2-4), 3.84 and 3.82 (each 3H, *s*, 2 $-OMe$), 3.33–2.26 (6H, *m*, H-2, H-3, H₂-5 and H₂-6), MS m/z 370 $[M]^+$, 219, 191, 151, 135, 77, 28.

Gadane (1a, 5 mg) was hydrogenated following the same procedure to the dihydro-compound 8 (4 mg), $C_{20}H_{18}O_6$, mp 132° (C_6H_6), IR $\nu_{max}^{KBr} cm^{-1}$ 1775 (γ -lactone), 1610, 1592, 1490 (aromatic $>C=C<$) and 940 ($-OCH_2O-$), 1H NMR δ 7.12–6.54

(6H, *m*, Ar-H), 5.98 and 5.95 (each 2H, *s*, 2 -OCH₂O-), 4.05 (2H, *br s*, H₂-4), 3.43-2.28 (6H, *m*, H-2, H-3, H₂-5 and H₂-6), MS *m/z* 354 [M]⁺, 203, 175, 135, 77, 28

Reactions of 2 and 1a with DDQ Jatrophan (2, 20 mg) was dissolved in dry C₆H₆ (20 ml), and DDQ (20 mg) added. The mixture was refluxed for 24 hr. The ppt formed was filtered off and the filtrate on concn afforded retrochinensin (11, 16 mg), which was purified by prep TLC on silica gel using C₆H₆-EtOAc (9/1), mp 232° (C₆H₆) (lit [8] 234-236°), UV λ_{max}^{EtOH} nm (log ε) 248 (4.60), 312 (3.96) and 348 (3.56), IR ν_{max}^{KBr} cm⁻¹ 1760 (γ-lactone), 1625, 1510, 1470 (aromatic >C=C<), 900 (-OCH₂O-); ¹H NMR (CDCl₃) δ 8.26 (1H, *s*, H-1), 7.39-6.83 (5H, *m*, Ar-H), 6.08 (2H, *s*, -OCH₂O-), 5.19 (2H, *s*, H₂-3α), 3.97 and 3.88 (each 3H, *s*, 2-OMe), MS *m/z* 364 ([M]⁺, base peak), 349, 335, 307, 249, 227, 163. The physical and spectral properties were indistinguishable from those reported for retrochinensin [8]. Direct comparison of the ¹H NMR spectrum of 11 with that of authentic retrochinensin confirmed the above identification.

Following the above procedure, 1a (10 mg) was also cyclized with DDQ to justicidin E (10, 7 mg), mp 265° (C₆H₆) (lit [7] 265-269°), IR ν_{max}^{KBr} cm⁻¹ 1762 (γ-lactone), 1620, 1510 (aromatic >C=C<), 930 (-OCH₂O-), ¹H NMR (CDCl₃) δ 8.27 (1H, *s*, H-1), 7.62-6.84 (5H, *m*, Ar-H), 6.12 and 6.16 (each 2H, *s*, 2 -OCH₂O-), 5.18 (2H, *s*, H₂-3α), MS *m/z* 348 ([M]⁺, base peak), 319, 291, 261, 233, 205, 176. The physical and spectral properties of 10 were identical to those reported for justicidin E [7].

Reaction of 2 with osmic acid To a soln of jatrophan (2, 20 mg) dissolved in pyridine (0.5 ml), OsO₄ (20 mg) was added with stirring and the reaction continued for 24 hr. A mixture of NaHSO₃ (35 mg), H₂O (5 ml) and pyridine (0.5 ml) was added and the reaction mixture stirred for 3 hr. The product was extracted with CHCl₃. Concn of the organic layer afforded 12 (14 mg), C₂₁H₂₂O₈, mp 175° (EtOH), IR ν_{max}^{KBr} cm⁻¹ 3560 (-OH), 1765 (γ-lactone), 1605, 1590 (aromatic >C=C<) and 912 (-OCH₂O-), ¹H NMR (CDCl₃) δ 7.27-6.57 (6H, *m*, Ar-H), 6.04 (2H, *s*, -OCH₂O-), 4.95 (1H, *s*, H-6), 4.55-4.10 (2H, *m*, H₂-4), 3.92 and 3.88 (each 3H, *s*, 2-OMe), 3.49-2.78 (3H, *m*, H-3 and H₂-5), 2.63 (2H, *br s*, exchangeable with D₂O, 2-OH), MS *m/z*, 402, 251, 233, 205, 175, 151 (base peak), 28.

Cyclization of 12 to 13 Compound 12 (10 mg) was added to a mixture of HCl (12 M) and HOAc (1/25) and the mixture stirred for 12 hr. The cyclization product 13, C₂₁H₂₀O₇, was extracted with CHCl₃ and purified by crystallization from EtOH, mp 245°, yield 6.5 mg, IR ν_{max}^{KBr} cm⁻¹ 3520 (-OH), 1770 (γ-lactone), 1610, 1590 (aromatic >C=C<) and 910 (-OCH₂O-), ¹H NMR (CDCl₃) δ 7.22-6.78 (5H, *m*, Ar-H), 6.04 (2H, *s*, -OCH₂O-), 4.79-4.14 (2H, *m*, H₂-3α), 4.43 (1H, *s*, H-1), 3.85 and 3.92 (each 3H, *s*, 2-OMe), 3.57-2.76 (3H, *m*, H-3 and H₂-4), 2.58 (1H, *br s*,

exchangeable with D₂O, -OH), MS *m/z* 384, 366 (base peak), 351, 333, 289, 182, 28.

Aromatization of 13 To a suspension of 10% Pd-C (10 mg) in *p*-cymene (10 ml), 13 (5.5 mg) was added. The resulting suspension was heated under reflux with stirring for 2 hr under N₂. The mixture was cooled, filtered and the solvent removed *in vacuo*. The residue on crystallization from EtOH furnished justicidin B (14, 4 mg), mp 235° (lit [9] 235-238°), IR ν_{max}^{KBr} cm⁻¹ 1760 (γ-lactone), 1615 (aromatic >C=C<), 930 (-OCH₂O-), ¹H NMR (CDCl₃) δ 7.73-6.92 (6H, *m*, Ar-H), 6.08 (2H, *s*, -OCH₂O-), 5.38 (2H, *s*, H₂-3α), 3.83 and 4.06 (each 3H, *s*, 2-OMe), MS *m/z* 364 ([M]⁺, base peak), 335, 321, 307, 277, 163, 28. The physical and spectral properties were indistinguishable from those reported for justicidin B [9].

NBS treatment of 2 Jatrophan (2, 20 mg) and NBS (20 mg) in dry CCl₄ (10 ml) were refluxed for 2 hr. The ppt formed was removed by filtration. The filtrate was evapd and the residue crystallized from EtOH to justicidin B (14, 15 mg), mp 235°. The compound was identical to the aromatization product of 13 (mmp, co-TLC and superimposable IR spectra).

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